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## SOIL & CROP SCIENCES | RESEARCH ARTICLE

# Relieving dormancy and improving germination of Piquín chili pepper (*Capsicum annuum* var. *glabriusculum*) by priming techniques

Maria Fernanda Quintero C<sup>1\*</sup>, Oscar Guillen Castillo<sup>1</sup>, Pablo Delgado Sánchez<sup>1</sup>, José Marín-Sánchez<sup>1</sup>, Ana Isabel Guzmán<sup>2</sup>, Agustín Sánchez<sup>2</sup> and José Miguel Guzmán<sup>2\*</sup>

**Abstract:** The effects of different priming techniques were evaluated to improve the dormancy and germination of wild seeds of “Piquín” chili pepper (*Capsicum annuum* var. *glabriusculum*). Three experiments were designed for pre-sowing treatment of seeds: (a) chemical seeds digestion; (b) halopriming (with  $K^+$  or  $NH_4^+$  of  $NO_3^-$ ,  $SO_4^{2-}$  or  $Cl^-$ ) at different priming times (24, 48 or 72 h) and osmotic potential ( $-5$ ,  $-10$  or  $-15$  atm) and (c) previously selected halopriming ( $KNO_3$  and  $NH_4NO_3$ ) + Gibberellic acid ( $GA_3$ , at 100 or 200  $mg \cdot L^{-1}$ ) were tested. Digestion treatments did show any effect on seed germination. Recommended values of osmotic potential ( $\Psi_s$ ), to improve Piquín chili seed germination, must be between  $-10$  and  $-15$  atm ( $-1.0$  and  $-1.5$  MPa) and the priming time must be between 48 and 72 h. Priming techniques can considerably reduce Capsaicinoids content on seeds, improve dormancy, seed germination performance, and increase the rate and uniformity of seedling establishment.  $KNO_3$  and secondly  $GA_3$  treatments may improve rapid and uniform

### ABOUT THE AUTHORS

Maria Fernanda Quintero C, Oscar Guillen Castillo, Pablo Delgado Sánchez, José Marín-Sánchez, Ana Isabel Guzmán, Agustín Sánchez and José Miguel Guzmán The paper is the result of a multidisciplinary work between the research groups: UASLP-CA-236 (UASLP) and RNM 151 PAIDI-UAL (CEI A3- UAL). It is a part of a master's thesis in agricultural production. Dra Quintero (UASLP-CA-236) and Dr Guzmán (RNM 151 PAIDI-UAL) are the main researchers, they carried out the experimental design, methodology and the final writing of work. Dr Delgado and Dr. Sánchez organized laboratory work and provided additional resources for their development. Postgraduate students Guillen and Guzmán, carried out the experiments and laboratory tasks, collaborated in information treatment writing of the paper. The work constitutes the preliminary stage of a broad project on “contribution to the domestication and production of Chile Piquín (*Capsicum annuum* L. var. *Glabriusculum*) in Mexico“. The project has been recently positively evaluated by the National Council of Science and Technology of Mexico, subject to budget availability.

### PUBLIC INTEREST STATEMENT

Piquín chili pepper has an appreciable culinary tradition in Mexico and the United States. Currently, wild populations of this specie have decreased significantly due to anthropogenic pressure and inadequate management of natural resources. To reduce this pressure, it is necessary to have techniques that allow to regeneration of wild populations and to improved cultivar's crop. The first problem to solve is the improvement of its low germination and to obtain seeds that acclimatize to both situations. Priming is recognized a technique that improves germination. The work aims to improve wild seeds germinability and reduce seedlings mortality. Priming with osmotically active fertilizer and gibberellic acid solutions has been effective for these purposes. Future research will aim, to maintain biodiversity, by adapting the technique to wild seeds for high risk ecotypes regeneration, and on the other hand to reduce the pressure on wild populations, selecting cultivars with greater domestication possibilities and the study of the proper crop's conditions.

germination and seedling emergence. The results provide basic information to develop guidelines for commercial establishment of Piquín pepper crops.

**Subjects: Agriculture & Environmental Sciences; Plant Reproduction; Agronomy; Environment & Resources; Conservation Environment Studies; Fruit & Vegetables**

**Keywords: domestication; seed; capsaicinoids; halopriming; gibberellic acid**

## 1. Introduction

Chili “Piquín” or “chiltepín”, [*Capsicum annuum* var. *glabriusculum* (Dunal) Heiser & Pickersgill]; syn. *C. annuum* var. *aviculare* (Dierb.) D’Arcy & Eshbaugh], is distributed from Colombia, Central America, and Mexico to the southwestern United States. The natural populations of “chiltepín” are considered an important genetic resource for pepper crop improvement (Hayano, Gamez, & Medina, 2016). This species is of great significance in the culture and identity of indigenous peoples of Mexico who usually harvested its fruits of wild plants (Guerrero Velázquez, 2015). The heat of chili pepper is due to the accumulation of capsaicinoids, a group of related alkaloids unique to *Capsicum*. Capsaicinoids are produced in the fruit placenta and transferred to the seeds during fruit maturation (Barchenger & Bosland, 2016). In highland regions where it occurs, is an important part of the local economy, especially in the time of harvest, generating employment and income for rural communities. This activity might threaten the genetic diversity in this species, affecting habitat degradation of natural populations of wild pepper. This problem could be solved by limiting the collection of wild populations and increasing their cultivation as a crop, in turn generating economic resources derived from this activity (Kim, 2016). While there is basic information that allows for developing guidelines for its cultivation, more research, related to germination, stand establishment and crop development and productivity, is necessary to develop commercial Piquín pepper crops.

Domestication of Piquín pepper plants have not been fully developed because problems are encountered related to low and erratic seed germination, morphologic and genetic variability, and limited environmental physiology information (Rodriguez-Urbe, Hernandez, Kilcrease, Walker, & O’Connell, 2014). Some authors suggest that germination of its seeds is restricted by physiological dormancy and is achieved after passing through the digestive tract of certain birds. Seeds of many species remain viable after passing through the digestive tracts of animals, with varying effects on germination (Cano-Vazquez et al., 2015). Seed dormancy is generally an undesirable characteristic in agricultural crops, where rapid germination and growth are required. Extensive domestication and breeding of crop species have ostensibly removed most dormancy mechanisms present in the seeds of their wild ancestors. Studies have reported a myriad of methods to break seed dormancy, including chemical, mechanical, thermal, and hormonal seed treatments (Paparella et al., 2015).

The beneficial effects of priming on the vigour, germination of seeds and establishment of the seedlings is known since the times of Pliny the elder (A.D. 23–79) (Gaius, 1949). Seed priming is a pre-sowing treatment that involves the controlled hydration of seeds, sufficient to allow pre-germinative metabolic events to take place but insufficient to allow primary root protrusion through the seed coat (Dutta, 2018; Paparella et al., 2015). It also involves complex physiological and biochemical process which offers an effective means to improve seed quality, seed germination and vigour (Bose, Kumar, Singhal, & Mondal, 2018; Siri, Vichitphan, Kaewnaree, Vichitphan, & Klanrit, 2013). Priming treatments are widely applied by seed companies to increase the germination rate and uniformity of seedling establishment of commercial vegetable and flower seeds. The benefits, associated with certain physiological, biochemical, cellular and molecular changes, include rapid, uniform and increased germination, improved seedling vigour and growth under a broad range of environments resulting in better stand establishment (Demir & Mavi, 2004). Different priming treatments can be effectively employed to prime many hot pepper seeds at one time (Ozbay, 2018; Paparella et al., 2015). Halopriming can affect osmoregulation in seeds by the active uptake of inorganic ions, promoting  $K^+$  and  $Ca^{2+}$

absorption and decreasing  $\text{Na}^+$  and  $\text{Cl}^-$  accumulation. Potassium plays an important role in balancing membrane potential and turgor, activating enzymes, and regulating osmotic pressure in cells (Ibrahim, 2016). Some authors hypothesized that capsaicinoids could have some allelopathic effect on pepper seed germination (Barchenger & Bosland, 2016). Capsaicinoids are a well-established allelochemical and has been shown to reduce root and shoot growth or suppress germination in several plant species (Kato-Noguchi & Tanaka, 2004). The effects of incorporating plant growth regulators into the priming solution have also been indicated to improve the germination and the growth of pepper seedlings (García, Montes, Rangel, García, & Mendoza, 2010; Quintero et al., 2017), and other vegetables.

The objective of this study was to evaluate both the response rate of wild seeds of Chili Piquín (*Capsicum annum* var. *glabriusculum*) to break dormancy and improve germination rate through seed priming and halopriming integrated with gibberellic acid ( $\text{GA}_3$ ) treatments. This information is needed to help in the development of sound and reliable guidelines for seedling production of Piquín pepper and contribute to its domestication.

## 2. Materials and methods

### 2.1. Plant materials

Fruit of Chili Piquín were collected from different wild populations in the States of Tamaulipas and San Luis Potosí, in North-eastern Mexico. Seed extraction was carried out manually, macerating fruits of each wild populations and dipping them in water to separate the pure seed from impurities. Seeds from different wild population were disinfected, as separate seed lot, in 1% sodium hypochlorite solution for 15 min to eliminate seed borne microorganisms. After disinfection, seeds were dried on shade for two weeks, then stored for three month, at laboratory hot and dry conditions (25°C, 50% RH), before giving the different treatments (Aloui, Souguir, & Hannachi, 2014).

### 2.2. Seed treatments

To achieve the proposed objective, a series of three consecutive experiments were designed for pre-sowing treatment of seeds. Each series of experiments was three times replicated for each stored seed lot. Following every treatment all seeds were rinsed under running tap water for 3 min and then with distilled deionized water for 1 min. After rinsing, seeds were surface dried by placing them between paper towels for 30 min. at room temperature. The seeds were then slowly dried at 25°C for 2 days until they reached their original moisture content (7–9%) and stored until capsaicinoids content determinations and germination test were carried out (Aloui et al., 2014). Untreated seeds were used as control and subjected to the same disinfection, rinsing and drying conditions.

#### 2.2.1. Digestion treatments

To simulate the effect of the digestive tract of birds on breaking dormancy on Piquín chili seeds, a group of seeds were subjected to a chemical wet digestion process using HCl and  $\text{H}_2\text{O}_2$ . Seeds were dipped in 0.2 N HCl for 5 min., and rinsed with dd $\text{H}_2\text{O}$  for 2 min. Subsequently were oxidized with 0.5 N hydrogen peroxide for 5 min and newly rinsed with dd $\text{H}_2\text{O}$  for 2 min. These redox sequences were repeated three times alternating both treatments (Jaganathan, Yule, & Liu, 2016).

#### 2.2.2. Priming treatments

Factorial halopriming was accomplished by imbibing 5 g of seed at 25°C in darkness for (24, 48 or 72 h) under an aerated solution of ( $\text{KNO}_3$ ,  $\text{K}_2\text{SO}_4$ ,  $\text{NH}_4\text{NO}_3$ , KCl,  $(\text{NH}_4)_2\text{SO}_4$  or  $\text{NH}_4\text{Cl}$ ) at  $-5$ ,  $-10$  or  $-15$  atm ( $-0.5$ ;  $-1.0$  or  $-1.5$  MPa respectively) of osmotic potential ( $\Psi$ s) to prevent seeds from entering the phase III of hydration (growth) (Aloui et al., 2014). Solutions were prepared by dissolving different salts in 250 ml Erlenmeyer glasses containing 100 mL of distilled water (Marín Sánchez., Mejía Contreras, Hernández Livera, Peña Lomeli, & Carballo Carballo, 2007). Untreated seeds were also used as control.

### 2.2.3. Priming integrated with gibberellic acid treatments

Priming, integrated with GA<sub>3</sub> treatment, was performed using two of the priming treatments [KNO<sub>3</sub> (-15 atm) and NH<sub>4</sub>NO<sub>3</sub> (-10 atm)], which further increased the germination parameters of the previous experiments. These priming treatments were supplemented with gibberellic acid (GA<sub>3</sub>) at 100 or 200 mg·L<sup>-1</sup>. Both controls (unprimed and without GA<sub>3</sub>) were used as absolute and relative control respectively. Indices were calculated referring to absolute control (untreated seeds) and to their respective relative control (priming treatments) and these denoted with the subscript.

### 2.3. Capsaicinoids determination

To test whether seeds capsaicinoids could be a contributor to seed germination, capsaicinoids content was determined on all seeds (primed and untreated) after treatments. Five-gram whole dry seeds were ground with a home blender for 3 min and then a fivefold volume of acetone was added, respectively, to the extract at 50°C for 1 h in triplicate. Centrifuged supernatant was taken for colorimetric analysis, following the methods proposed by Wang-Kyun, Hee-Woong, Geun-Dong, and Hae-Ik (2017).

### 2.4. Germination tests

These were carried out in darkness in a temperature-controlled incubator held at 25 ± 0.5°C and 100% relative humidity (Guerrero Velázquez, 2015). Seeds were placed on two layers of filter paper moistened with 3 mL of distilled water in covered 10 cm petri dishes. Germination values were recorded daily for 28 days to establish statistical data. From the total number of germinated seeds, final germination percentage (FGP) was calculated. For ungerminated seeds, tetrazolium chloride tests were conducted to differentiate between dormant and dead seeds (ISTA, 2017). Final latent percentage (FLP) and final mortality percentage (FMP) of seeds were calculated accordingly.

Primary root protrusion to 1 mm was scored as germinated seed. To evaluate root growth, a network of fiberglass of 1 mm<sup>2</sup> was placed under seeds. Primary root length (PRL) was measured in mm. Development germination index (DGI) allows to quantify effects (including FGP and PRL) of treatments (t) respect to control (o) on germination development. DGI was calculated by Zucconi tests (Selim, Zayed, & Atta, 2012) by following the formula:

$$[DGI = 100 \cdot (FGP(t)/FGP(o)) \cdot (PRL(t)/PRL(o))].$$

Days to 50% of FGP (T<sub>50</sub>) and days between 10% and 90% of FGP (G<sub>10-90</sub>) were also calculated. T<sub>50</sub> is an inverse measure of mean germination rate, while G<sub>10-90</sub> is an estimate of germination uniformity (GU) or germination synchrony. To contrast the behaviour of treatments(t) to control(o), these parameters were transformed in their respective indices, according to the following formulas: Rate germination index [RGI = 100·(T50(t)/T50(o))]; synchrony germination index [SGI = 100·(G<sub>10-90</sub>(t)/G<sub>10-90</sub>(o))].

After germination testing, germinated seeds were transplanted to conventional seedling trays inside a greenhouse to evaluate the number of abnormal seedlings generated by each treatment. Abnormal seedling percentage (ASP) and its corresponding abnormality seedling index [ASI = 100·(ASP(t)/ASP(o))], were calculated from abnormal plantlets.

### 2.5. Experimental design and statistical analysis

Treatments were arranged in completely randomized factorial design with four replications of 25 seeds. Data were subjected to multifactorial ANOVA. Mean separation was performed by Fisher's least significant difference (LSD<sub>0.05</sub>) test if F test was significant at  $p \leq 0.05$  (\*).

## 3. Results and discussion

Capsaicinoids contents, germination parameters, primary root growth and transplant abnormality for each seed treatment are shown in Tables 1–3 respectively. No differences were found between

**Table 1. Average values, ANOVA significance and LSD<sub>0.05</sub> values of *Capsicum annuum* var. *glabriusculum/aviculare* seeds and seedless, germinated in darkness at 25 °C following digestion treatments**

	CC ( $\mu\text{g}\cdot\text{g}^{-1}$ )	FGP (%)	PGI	FLP (%)	LGI	FMP (%)	MGI	G <sub>50</sub> (d)	RGI	G <sub>10-90</sub> (d)	SGI	PRL (mm)	DGI	ASP (%)	ASI
Significance.	NS	NS	*	NS	*	NS	*	NS	*	NS	*	NS	**	NS	*
Control seeds	973	8.1	100b	46.3	100b	43.9	100a	25.2	100a	14.1	100b	25.4	100b	15.1	100
Digested seeds	1007	7.7	92a	45.0	92a	46.4	116b	23.5	111b	14.3	91a	17.5	66a	15.9	114
LSD <sub>0.05</sub>	260	0.66	4.14	3.63	3.62	4.01	6.68	2.04	5.91	1.18	4.71	10.98	13.9	1.20	9.84

Capsaicinoids Content (CC); Final Latent Percentage (FLP); Final Mortality Percentage (FMP); Final Germination Percentage (FGP); Primary Root Length (PRL); Development Germination Index (DGI); Days to 50% of FGP (T50); Rate Germination Index (RGI); Days between 10% and 90% of FGP (G<sub>10-90</sub>); Synchrony Germination Index (SGI); Abnormal Seedling Percentage (ASP); Abnormality Seedling Index (ASI).

Means within the same column followed by the same letter are not different at  $p \leq 0.05$  per Fisher's least significant difference (LSD) test.

NS, \* Nonsignificant or significant at  $p \leq 0.05$ .

**Table 2. Average values, ANOVA significance and LSD<sub>0.05</sub> values of *Capsicum annuum* var. *glabriusculum* seeds and seedling, germinated in darkness at 25°C following priming (Pr) treatments**

	CC ( $\mu\text{g}\cdot\text{g}^{-1}$ )	FLP (%)	FMP (%)	FGP (%)	PRL (mm)	DGI	T <sub>50</sub> (d)	G <sub>10-90</sub> (d)	ASP (%)
Priming salt	*	*	*	*	*	*	*	*	*
Control	957d	44.7c	48.8c	7.9a	18.3c	100a	25.5e	14.6c	14.8c
NH <sub>4</sub> Cl	638c	39.2ab	26.2b	36.5c	17.1abc	421bc	22.4d	21.1f	12.9bc
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	469b	39.9abc	28.0b	32.4b	16.8abc	366bc	22.1cd	22.1g	11.4b
NH <sub>4</sub> NO <sub>3</sub>	319a	43.2bc	11.9a	45.0d	16.8abc	501de	21.0b	12.0b	8.6a
KCl	579c	39.7ab	28.2b	32.7bc	16.0a	360b	21.1bc	17.1d	12.8bc
K <sub>2</sub> SO <sub>4</sub>	441b	36.8a	27.2b	35.3bc	17.6abc	437cd	20.8b	18.9e	11.7b
KNO <sub>3</sub>	309a	40.2abc	13.5a	46.1d	17.8bc	565e	19.0a	11.0a	10.1ab
Priming time (h)	*	*	*	*	*	*	*	*	*
0	957c	44.7c	48.8d	7.9a	18.3c	100a	25.5d	14.6a	14.8c
24	607b	49.7d	16.9a	33.7b	15.2a	353b	22.7c	18.9c	13.8c
48	420a	38.3b	23.7b	38.5c	16.6b	433c	21.3b	17.2b	11.6b
72	350a	31.5a	26.8c	41.8d	19.2c	538d	19.2a	15.0a	8.4a
Priming $\Psi_s$ (atm)	*	*	*	*	*	*	*	*	*
0	957c	44.7c	48.8c	7.9a	18.3b	100a	25.5d	14.6a	14.8c
-5	555b	46.2c	20.9a	34.0b	15.5a	368b	22.2c	18.2c	14.3c
-10	395a	38.7b	21.8ab	39.7c	17.3b	462c	21.2b	17.0bc	8.8a
-15	428a	34.5a	24.8b	40.3c	18.1b	496c	19.8a	16.0ab	10.8b
Ps × Pt	*	*	*	*	*	*	*	*	*
Ps × P <sub>ψ</sub>	*	*	*	*	*	*	NS	*	*
Pt × P <sub>ψ</sub>	*	*	*	*	*	*	NS	NS	*

Capsaicinoids Content (CC); Final Latent Percentage (FLP); Final Mortality Percentage (FMP); Final Germination Percentage (FGP); Primary Root Length (PRL); Development Germination Index (DGI); Days to 50% of FGP (T<sub>50</sub>); Days between 10% and 90% of FGP (G<sub>10-90</sub>); Abnormal Seedling Percentage (ASP).

\* Significant differences at  $p \leq 0.05$ . Means within the same column followed by the same letter are not different at  $p \leq 0.05$  per Fisher's least significant difference test.

**Table 3. Average values, ANOVA significance and LSD<sub>0.05</sub> values of *Capsicum annuum* var. *glabriusculum* seeds and seedling, germinated in darkness at 25 °C following pre-sowing with gibberellic acid treatments and priming integrated with gibberellic acid (GA<sub>3</sub>) treatments**

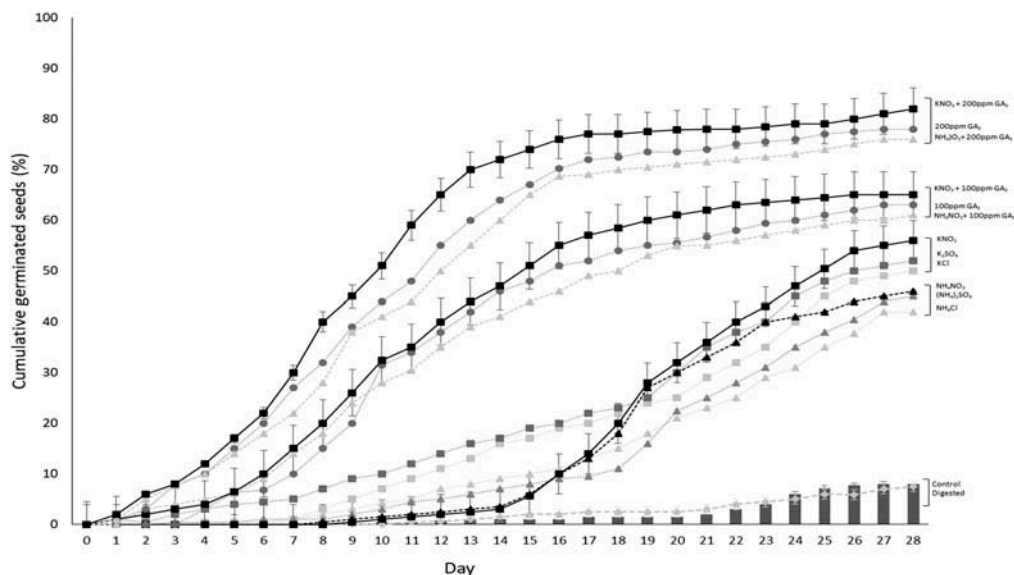
Treatment	CC (µg·g <sup>-1</sup> )	FLP (%)	FMP (%)	FGP (%)	PRL (mm)	DGI	T <sub>50</sub> (d)	G <sub>10-90</sub> (d)	ASP (%)
Control	824e	47.0d	45.4f	7.6a	21.4c	100a	26.7f	14.9de	32.0f
+100mg·L <sup>-1</sup> GA <sub>3</sub>	491d	11.9b	27.1e	60.1d	13.4a	545b	10.8bc	15.6e	21.4e
+200mg·L <sup>-1</sup> GA <sub>3</sub>	384c	7.1a	16.9c	76.0f	13.7a	674c	8.2a	8.8a	15.7d
NH <sub>4</sub> NO <sub>3</sub> (-10 atm)	461d	43.4c	8.2ab	48.4b	18.0b	579b	22.8e	12.9c	6.9a
+100 mg·L <sup>-1</sup> GA <sub>3</sub>	201b	12.0b	23.4d	64.6e	30.9d	1331d	11.3c	15.5e	20.6e
+200 mg·L <sup>-1</sup> GA <sub>3</sub>	74a	12.2b	16.4c	76.9f	39.8e	2036e	8.0a	8.0a	7.0a
KNO <sub>3</sub> (-15 atm)	402c	42.4c	5.4a	52.3c	19.1b	665c	18.4d	10.2b	11.2b
+100 mg·L <sup>-1</sup> GA <sub>3</sub>	209b	6.7a	21.6d	66.3e	28.9d	1276d	10.1b	14.2d	13.2c
+200 mg·L <sup>-1</sup> GA <sub>3</sub>	72a	6.8a	11.5b	81.8g	38.4e	2096e	8.1a	8.1a	11.1b
NO <sub>3</sub> × GA	*	*	*	*	*	*	NS	*	*

Capsaicinoids Content (CC); Final Latent Percentage (FLP); Final Mortality Percentage (FMP); Final Germination Percentage (FGP); Primary Root Length (PRL); Development Germination Index (DGI); Days to 50% of FGP (T<sub>50</sub>); Days between 10% and 90% of FGP (G<sub>10-90</sub>); Abnormal Seedling Percentage (ASP).

\* Significant differences at  $p \leq 0.05$ . Means within the same column followed by the same letter are not different at  $p \leq 0.05$  per Fisher's least significant difference (LSD) test.



**Figure 1. Cumulative germination percentage of Piquín pepper after seeds treatments (digested or primed) monitored for 28 days. Error bars are presented only for control, digested and KNO<sub>3</sub> treatments.**



seeds lot or replications. The average daily percent germination values for treatments and control over a 28-day germination period are shown in Figure 1.

### 3.1. Digestion treatments

Table 1 shows germination parameters of seeds digested with HCl and H<sub>2</sub>O<sub>2</sub>. Average values show no significant difference for CC, FLP, FMP, FGP, PRL, T<sub>50</sub>, G<sub>10-90</sub>, or ASP, while significant differences for DGI, RGI, SGI and ASI indices were found, indicating that these indices, are more sensitive to detect the treatment effects referred to control than the proper parameters. The chemical digestion of Piquín pepper seeds does not affect capsaicinoids content (CC) on seeds. The lower FGP and PRL of digested seeds lead to a strong reduction on DGI (-33%) indicating a marked detrimental effect on germination development. Digestive treatments only increase mean germination rate (+11% RGI) and could contribute to break dormancy or latency reducing FLP (Table 1), but also reduces synchrony (-9% SGI), increases FMP, does not improve FGP, and strongly worsen early developmental stage of seedling and abnormality of transplants (+9% ASI).

### 3.2. Priming treatments

Average values of germination parameters and DGI are presented on Table 2. Significant differences were found in all factors of priming treatment (salt, time and Ψs) for all parameters and indices. Different behaviour was observed for different salts, showing differences between K<sup>+</sup>- and NH<sub>4</sub><sup>+</sup>- salts on FGP (Figure 1) and between NO<sub>3</sub><sup>-</sup> and SO<sub>4</sub><sup>2-</sup> or Cl<sup>-</sup> on synchrony (Table 2). All treatment reduces capsaicinoids content on primed seeds. Highest CC reduction were obtained (Table 2) on seeds primed with NO<sub>3</sub><sup>-</sup> salts (more than SO<sub>4</sub><sup>2-</sup> or Cl<sup>-</sup>) and at -10 or -15 atm (more than -5), for 48 or 72 h (more than 24).

FGP was increased four to five times for Cl<sup>-</sup> and SO<sub>4</sub><sup>2-</sup>. NO<sub>3</sub><sup>-</sup> salts (of NH<sub>4</sub><sup>+</sup> or K<sup>+</sup>) increased FGP (6 times) and reduced to 25% seeds mortality (Table 2). A higher final percent of germinated seeds was also obtained for K<sup>+</sup> rather than NH<sub>4</sub><sup>+</sup> containing salts (Figure 1). Highest FGP (together with low effect on PRL reduction) of NO<sub>3</sub><sup>-</sup> primed seeds lead to a strong increase on DGI, indicating a clear improvement on germinative process. DGI increases three to four times for Cl<sup>-</sup> and SO<sub>4</sub><sup>2-</sup> and by five times for NO<sub>3</sub><sup>-</sup>. KNO<sub>3</sub> increased DGI reducing T<sub>50</sub> and G<sub>10-90</sub> more than NH<sub>4</sub>NO<sub>3</sub>, whereas NH<sub>4</sub>NO<sub>3</sub> reduced ASP more than KNO<sub>3</sub>. An incremental effect was observed for priming time and Ψs on FGP, DGI and T<sub>50</sub>. Increments on germination rate were 6-12% higher using K<sup>+</sup> than NH<sub>4</sub><sup>+</sup> containing salts (Table 2). Latent seeds were only significantly reduced for K<sub>2</sub>SO<sub>4</sub> or NH<sub>4</sub>

Cl salts at  $-10$  or  $-15$  atm for 48 or 72 h. Radicle length was only significantly reduced on KCl primed seeds under  $\Psi_s -5$  atm for 24 or 48 h.

A differential effect was observed on germination synchrony for different factors. Germination synchrony increases on nitrate primed seeds (lowest  $G_{10-90}$ ), whereas was reduced on seeds primed with sulphate or chloride. Priming times shorter than 72 h, or lower than  $-10$  atm of  $\Psi_s$  on priming solution, reduces synchrony (highest  $G_{10-90}$  on Table 2). Figure 1 shows the average percentage germination values over time for all priming and digestion treatments. A different behaviour appears on the germination process for each treatment during 28 days of germination. Germination synchronies ( $G_{10-90}$  on Table 2) were expanded by  $\text{Cl}^-$  and  $\text{SO}_4^{2-}$  whereas reduced by  $\text{NO}_3^-$ . Seeds primed with nitrate containing salts clearly increases germination synchrony and mean germination speed, but the effect not appears to be responsible for breaking of dormancy. Seeds latency (FLP) could probably be improved by including  $\text{GA}_3$  in priming solutions (Figure 1).

Abnormality of plantlets was reduced as priming time increases and was lower for  $\Psi_s -10$  atm. ASP reduced 38% for  $\text{Cl}^-$ , 62% for  $\text{SO}_4^{2-}$  and 70% for  $\text{NO}_3^-$ . Graphic analysis of interactions indicated that 72 h priming treatments with  $\text{NH}_4\text{NO}_3$  ( $-15$  atm) and  $\text{KNO}_3$  ( $-10$  atm) are optimum regarding the improvement of FGP, PRL, DGI and ASP.

### 3.3. Priming integrated with gibberellic acid treatments

Average values of germination parameters and indices are presented on Table 3. All treatment significantly reduces capsaicinoids content on primed seeds. Highest CC reduction were obtained on seeds primed with  $\text{NO}_3^-$  salts and at  $200 \text{ mg}\cdot\text{L}^{-1}$  of  $\text{AG}_3$ . Combined effects of nitrate priming and  $\text{AG}_3$  reduces initial capsaicinoids contents to 10%. A high correlation between DGI and CC were found ( $\text{DGI} = (60.07 - 1.71 \cdot \sqrt{\text{CC}})^2$ ;  $R^2 = 96.91\%$ ).

Pre-sowing with gibberellic acid treatments (Control +100 or +200  $\text{mg}\cdot\text{L}^{-1}$   $\text{GA}_3$ ) also shows (Figure 1) a positive effect on germination respective to absolute control for all evaluated parameters (Table 3), except PRL (100 and 200  $\text{mg}\cdot\text{L}^{-1}$ ) and  $G_{10-90}$  (100  $\text{mg}\cdot\text{L}^{-1}$ ).

$\text{GA}_3$  significantly reduces latency (FLP) in Piquín chili seeds (Table 3) referred to the absolute control and maintains this effect when it is added to priming solutions (Figure 1). The addition of  $\text{GA}_3$  (at 100 or 200  $\text{mg}\cdot\text{L}^{-1}$ ) activates dormant seeds to a rate between 73 and 84% respectively. This latency inhibition causes an increase in FGP of between 30 and 60%. However,  $\text{GA}_3$  additions to priming solutions increases FMP respect to controls.

$\text{GA}_3$  significantly increases mean germination rate (reducing  $T_{50}$  on Table 3) compared to absolute or relative controls. At 200  $\text{mg}\cdot\text{L}^{-1}$  this  $T_{50}$  decrease by 2.5 times. However, the effect of  $\text{GA}_3$  on synchrony is different. While additions of 100  $\text{mg}\cdot\text{L}^{-1}$  has no effect, addition of 200  $\text{mg}\cdot\text{L}^{-1}$  double the synchrony, reducing germination time ( $G_{10-90}$ ) from 15 to 8 days. These synergic effects of the addition of  $\text{GA}_3$  to priming solutions is clearly show for germination percentages on Figure 1. Conversely, 200  $\text{mg}\cdot\text{L}^{-1}$   $\text{GA}_3$  has no effect on ASP, while 100  $\text{mg}\cdot\text{L}^{-1}$   $\text{GA}_3$  significantly increases the presence of abnormal seedlings in primed seeds.

Gibberellic acid applied alone, significantly reduces the length of the primary root with respect to the absolute control. However, the integrated priming treatment with  $\text{GA}_3$ , practically duplicate PRL for  $\text{GA}_3$  (200  $\text{mg}\cdot\text{L}^{-1}$ ) and increases it by between 50 and 70% for  $\text{GA}_3$  (100  $\text{mg}\cdot\text{L}^{-1}$ ). These increases in PRL together with the originated in FGP lead to double or triple values of DGI (associated with  $\text{GA}_3$ ) compared to their respective relative controls. On the other hand, the reduction in PRL (associated with the application of  $\text{GA}_3$ ) regarding the absolute control, neutralizes the positive impact generated on FGP and originates DGI increases, on relative control, like those produced by the halopriming without  $\text{GA}_3$ .

### **3.4. Digestion treatment**

While some authors argue that Piquín chili seed germination increases after passage through the digestive tract of birds (Arcia González, 1985; Bañuelos, Salido, & Gardea, 2008), evidence of this fact has not been provided (Araiza Lizarde, Araiza Lizarde, & Martínez Martínez, 2011). Digestive treatments could contribute to breaking dormancy, increasing mean germination speed, but do not improve germination percentage or synchrony and strongly worsen early developmental stage of seedlings. The positive effects on germination related to birds appear to be more associated to the dispersal and deposition of seeds in favourable environments that on stimulate further germination. Digestion treatments have not shown any positive effect on the germination of Piquín chili seeds of. Authors have presented both similar results (Reid & Armesto, 2011), and have also found large differences (Guerrero Velázquez, 2015; Jaganathan et al., 2016) in the behaviour of different accessions of plants.

### **3.5. Priming treatments**

Priming has been proposed as a mechanism of invocation of different stress tolerance of germinating seeds. Seed priming treatments have been applied to various crops under saline conditions (Ruttanaruangboworn, Chanprasert, Tobunluepop, & Onwimol, 2017). Some authors find that a specific ion or salt is not essential to priming pepper seed (Smith & Cobb, 1991), or other horticultural crop (Siri, Vichitphan, Kaewnaree, Vichitphan, & Klanrit, 2013). Nitrate enhanced germination and seedling establishment rates under adverse conditions, of onion, tomato, asparagus, melon, watermelon, husk tomato and pepper (Marín Sánchez, Rivas Jacobo, Flores Cano, Rojas Velazquez, & Jarquin Gálvez, 2013). Our results also indicate that nitrate-containing salts are more efficient than nitrate-free salts on promoting germination of primed seeds (except breaking dormancy). In addition, the effects of priming with  $\text{KNO}_3$  seem to be more positive than  $\text{NO}_4\text{NO}_3$  on main germination and seedling establishment parameters (except for seed mortality and seedling abnormality). Seed priming stimulates pre-germinative metabolic processes and prepares seeds for primary root protrusion. Priming increases antioxidant system activity and repair membranes, moreover the reduction of capsaicinoids on seeds during priming, could contribute to break dormancy and stimulate germinative process. These changes promote seed vigour during germination and emergence (Ibrahim, 2016).

Time-course experiments show that effective priming is strongly dependent on both the osmotic potential of the priming solution and the duration of the treatment to avoid “over priming” (Smith & Cobb, 1991). Accordingly, the recommended values of osmotic potential to improve germination of Piquín chili seed must be between  $-10$  and  $-15$  atm ( $-1.0$  and  $-1.5$  MPa) and the treatment time must be between 48 and 72 h.

A small number of studies were developed on Piquín pepper germination. All of them are heavily dependent on the origin of seeds accessions and on genetic diversity and present conflicting results (Cano-Vázquez et al., 2015; Guerrero Velázquez, 2015; Rodríguez-Uribe et al., 2014). Authors do not find positive effects of  $\text{KNO}_3$  priming, whereas only positive effects with  $\text{GA}_3$  at extremely high doses ( $5000 \text{ mg}\cdot\text{L}^{-1}$ ) were observed. However, none of these studies combine priming with  $\text{GA}_3$  at low doses. The undesirable observed effects of seed latency mean germination time ( $T_{50}$ ) and synchrony ( $G_{10-90}$ ) could be improved by including gibberellic acid ( $\text{GA}_3$ ) in osmotically active priming solutions (as shown in Figure 1).

### **3.6. Priming integrated with gibberellic acid treatments**

Halopriming with the addition of plant growth regulators may be an effective way to shorten emergence time and increase stand establishment in watermelon and pepper at low temperatures (Korkmaz, 2005). Halo-priming using  $\text{KNO}_3$  or a growth regulator like  $\text{GA}_3$  improves germination rate and reduces the mean germination time (Tzortzakis, 2009).

The integration of priming with  $\text{GA}_3$  was effective in improving germination and establishment of pepper and tomato seeds. Priming, during which germination is suspended, provides a unique way to

rapid and efficiently digest the endosperm by GA-induced enzymes and reduce the mechanical restraints of endosperm thus providing energy to start and sustain embryo growth. Studies of genetics and physiology have shown the important roles of the plant hormones such as abscisic acid and gibberellin in the regulation of seed dormancy and germination (Koorneef, Bentsink, & Hilhorst, 2002).

Considerable improvements on germination performance (rate and uniformity) and seedlings establishment are shown for KNO<sub>3</sub> and GA<sub>3</sub> treatments, in agreement with Tzortzakis (2009). The lowest values of capsaicinoids found on KNO<sub>3</sub> + 200 mg·L<sup>-1</sup> GA<sub>3</sub> primed seeds could reduce their allelopathic effect on germination. Since high concentrations of capsaicin inhibit the germination of chili seeds (Barchenger & Bosland, 2016), the positive effects on germination may be due to the elimination of these as germination inhibitors (Cano-Vazquez et al., 2015).

#### 4. Conclusions

This study showed that it is possible to relieve dormancy and improve germination performance on Piquín chili seeds by priming techniques. Wild Piquín chili seed primed with KNO<sub>3</sub> (-10 atm; 72 h) + GA<sub>3</sub> (200 mg·L<sup>-1</sup>) improved germination (percentages and synchronicities) and reduced the start-up time of germination and seedling abnormality. Moreover, the results of current study provide essential information needed for the development of guidelines for the domestication and cultivation of Piquín chili plants.

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#### Author contributions

Conceived and designed the experiments: MFQ and MG. Performed the experiments: OG, AGC and AGS. Analysed the data: MFQ, PD, AGS and MG. Contributed reagents/materials/analysis tools: PD, and AGS. Wrote the paper: MFQ and MG

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