This is an Accepted Manuscript of an article published by Taylor & Francis in [Journal of Environmental Science and Health, Part A] on [November 2002], available at: https://doi.org/10.1081/ESE-120004524

INFLUENCE OF THERMAL REGIME OF SOIL ON THE SULFUR (S) AND SELENIUM (Se) CONCENTRATION IN POTATO PLANTS

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Key Words: Solanum tuberosum; Mulch; Polyethylene; Phytoremediation; Root-zone
 temperature

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19 ABSTRACT

20 Three consecutive years of field experiments were carried out to investigate the effect 21 of different root temperatures, induced by the application of mulches on the 22 concentration of sulfur (S) forms (organic-S, total-S and SO²₄) and Se in different organs of potato plants (roots, tubers, stems and leaves). Four different plastic covers were 23 24 used (T1: transparent polyethylene; T2: white polyethylene; T3: white and black 25 coextruded polyethylene, and T4: black polyethylene), using uncovered soil as control 26 (T0). The different treatments had a significant effect on mean root temperatures 27 (T0=16°C, T1=20°C, T2=23°C, T3=27°C and T4=30°C) and induced a significantly different 28 response in the S forms and Se concentration, showing the T3 treatment (27°C) the 29 greatest concentration of total S and organic S in the stems and leaflets. The Se reached 30 higher levels in the roots and tubers in T3. With regard to possibilities in 31 phytoremediation, it is necessary to control the thermal regime of the soil to optimize 32 the accumulation of elements.

33

34 INTRODUCTION

Root zone temperature strongly influences the growth and uptake of nutrients.^[1] The potato plants require optimal temperatures in the root zone for maximum growth and yield^[2] and one of the techniques used to increase root zone temperature is the application of polyethylene covers (mulch) of different colours and characteristics, which generate a favourable microenvironment (higher temperatures) in the root zone.^[3]

In its reduced form, sulfur (S) has an important function in growth and regulation
 of plant development,^[4] because of its essential role in the synthesis of amino acids,
 proteins and some secondary metabolites.^[5]

While selenium (Se) is not an essential plant nutrient,^[6] and exerts toxic effects in plants principally by interfering with sulfur (S) metabolism,^[7] this elements is essential for maintaining mammalian health. Benefits attributed to proper Se nutrition range 47 from immune system enhancement to cancer suppression.^[8] However, when consumed
48 in high quantities, Se can accumulate in tissues and become toxic.^[9]

The current problem of the pollution of agricultural soils and waters causes problems for human health which can be partially solved with the application of technology of phytoremediation.^[10] The objective of this technology is to eliminate contaminants for the environment by using plants.^[11]

53 The aim of the present work was to evaluate the effect of the different root zone 54 temperatures generated by the application of mulches on S status and Se concentrations 55 using field grown potato plants. The aim of the present work was to evaluate the effect 56 of the different root zone temperatures generated by the application of mulches on S 57 status and Se concentrations using field grown potato plants.

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59 MATERIALS AND METHODS

60 Crop Design

61 The experiment was conducted for three consecutive years (1993–1995) in the 62 field (Granada, Spain), using Solanum tuberosum L. var. Spunta, planted at the beginning 63 of March and the crop cycle was about 4 months. The climate was semiarid and the area 64 intensively used for agriculture. The soil used showed the following characteristics: sand 65 45.3%, silt 43.2%, and clay 11.2%, pH (H₂O 1:2.5) 8.6; electrical conductivity 1.10dSm⁻¹, 66 CaCO₃ 11.2%; total N (0.1%); P₂O₅ (58µgg⁻¹); K₂O (115µgg⁻¹); DTPA+TEA+CaCl₂ (pH 7.3) 67 extractable Se 24µgkg⁻¹. The characteristics of the irrigation water were: pH 7.6; E.C. 1.05dSm⁻¹; Cl 58mgL⁻¹; Na⁺ 25mgL⁻¹; K⁺ 4mgL⁻¹; H₂CO₃ 369mgL⁻¹, Se 1µgL⁻¹, SO⁻²₄ 90mgL⁻¹. 68

The experimental design was a factorial arrangement in a randomized complete block with 5 treatments replicated 4 times (20 plots). Each plot occupied an area of 78.4m⁻², with a planting density of 4.2 plants m⁻². Plants were spaced 30cm apart, with 80cm between rows. The soil temperature was measured at the 15-cm in depth, using probes (107 type) from Campbell Scientific TM. Root zone temperature was measured (6 measurements at 4-h intervals) every 3 days of the crop cycle.

The different treatments consisted of covering the soil surface of each plot with plastic mulches (polyethylene sheets), making a tight seal with the soil: transparent polyethylene (25mm in thickness, T1), white polyethylene (25mm in thickness, T2), coextruded black and white polyethylene (50mm in thickness, T3), and black polyethylene (25mm in thickness, T4). Finally, no plastic was applied in the control treatment (T0).

The fertilization used was the same as is habitually applied by farmers in the zone. In the month of February in all three years, N (NH₄NO₃) and P and K (K₂HPO₄) were applied (27gm⁻²). Afterwards, at the end of the month of April, 25gm⁻² of NH₄NO₃ were applied. Fertigation was complemented with the following micronutrients: Fe: 0.5mgL⁻ 1; B: 0.1mgL⁻¹; Mn: 0.1mgL⁻¹; Zn: 0.075mgL⁻¹; Cu: 0.075mgL⁻¹ and Mo: 0.05mgL⁻¹. Iron was applied as FEEDDHA, B as H₃BO₃ and the remaining micronutrients as sulphates.

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88 Plant Sampling

The plant material (stems, leaves, roots and tubers) were sampled 6 times every two weeks, throughout the plant development for the three years of experiments. For each sampling, 10 plants were collected from each replicate per treatment. Leaf samples were taken only from plants with fully expanded leaves of the same size. Leaves were picked at about one third of the plant height from the plant apex. Roots, leaves, stems
and tubers were rinsed three times in distilled water after decontamination with nonionic detergent at 1%, then blotted on filter paper. Then a sample was dried in a forced
air oven at 70°C for 24h, ground in a wiley mill and then placed in plastic bags for the
further analyses.

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99 Plant Analysis

100 Se Determination:

For the assay of total Se concentration, oven-dried and pulverized plant material was digested with concentrated nitric acid and measurements were made using an atomic absorption spectrophotometer equipped with a graphite furnace.^[12] Reagent blanks for analysis were also prepared performing the entire extraction procedure but in the absence of the samples.

106

107 Sulfate Determination:

Sulfate (SO²⁻₄) was determined in aqueous extraction of 0.2g of dried ground material in 10mL of MILLIPORE-filtered water, shaking 120min at room temperature and then filtered with Whatman-n1 filter paper. SO²⁻₄ was determined from the aqueous extract obtained and measured by turbidimetry of the BaSO₄ maintained in suspension by means of tensioactive agent (gum arabic) according to Novozamsky and Van Eck.^[13]

114 Organic-S and Total-S Determination:

115 A 0.1g dry weight sub-sample was digested with nitric acid mineralization and 116 H_2O_2 .^[14] After dilution with deionized water, in the product, organic S was measured as 117 described previously for the sulphate,^[13] against a pattern curve. Total S were 118 recalculated from the sum of organic S and sulphate.

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120 Statistical Analyses

121 Analysis of variance was used to assess the significance of treatment means. 122 Significant differences according to the Duncan's Multiple Range Test (DMRT) are 123 indicated with different letters in the tables. Level of significance are represented by * 124 at p<0.05, ** at p<0.01, *** at p<0.001, and ns: not significant.

125

126 **RESULTS AND DISCUSSION**

127 Table 1 shows the mean root zone temperatures (RZT) generated under the 128 different treatments with the highest value in T4 (30°C), and the lowest in T0 (16°C). The 129 effect of the different mulches on root-zone temperatures were similar to those of Ham 130 et al.,^[15] who reported that black polyethylene (our T4), absorbs roughly 96% of the incoming radiation while reflecting very little, thus warms the soil.[16] The white 131 132 polyethylene covers (T2) induced a cooler soil temperature than did black covers (T4) 133 because the former reflected most wavelengths than transparent mulches (T1) do not 134 cause soil warming, presenting mean temperatures of 18–20C whereas the whiteb black 135 coextruded covers generated higher mean RZT (27°C in T3).

Table 1 also presents the results of the biomass (in a dry-weight basis) for the different organs of the potato plants. The dry mass was significantly affected by the RZT and showing for the roots, leaves and tubers, the highest values in T3 (27°C), and the lowest in T1 (20°C), the latter being lower than in T0 (16°C). On the contrary, in the stems, T1 reached the highest dry weight while T3 showed the lowest. Similarly to the results of Klock et al.,^[17] with tomato plants, the increase in total biomass was obtained in plants within the root-zone temperature ranging of 23–27°C in roots, tubers and leaflets, while outside this range (T0, T1 and T4), the dry weight fell with a lower dry weight accumulation (Table 1).

145 In relation to the effect of the different root-zone temperatures on Se in roots, 146 T3 reached the highest concentration surpassing T0 by 51%, while the lowest value was 147 found in T1 (Table 2). In tubers, the Se concentrations were below the limit of the 148 detection of the employed technique, whereas in stems T4 presented the highest Se 149 concentration (9% higher than in T0), and the lowest in T2. Finally, the highest and the 150 lowest Se concentrations in leaflets were recorded in T3 (60% higher than in T0) and T1 151 (22% lower than T0), respectively.

Root-zone temperature strongly influences the uptake of elements.^[1,18] In our experiments, the RZT treatments significantly increased the Se in the roots and leaflets in T3 and T4 with higher root temperatures (Table 2) increasing the root absorption and its transport to the aerial part. The failure to detect Se in the tubers was possibly due to the minor translocation of this element via the phloem,^[19] since edible plant parts (tubers) contain much less Se than do the inedible parts.^[20]

Non-significant differences were found between the treatments for SO^{2-4} concentration in roots (Table 3), whereas in tubers, the highest concentration was found in T1 (exceeding T0 by 56%) and the lowest in T2 and T3 (33% and 35% lower than T0, respectively). In stems, higher SO^{2-4} concentration were found in T0 and T1 whereas T2, T3 and T4 were significantly lower. Finally, T1 gave highest leaflets SO^{2-4} concentration (15% higher than in T0), and the lowest in T3 (10% less than in T0).

Since SO²⁻₄ is absorbed in low quantities,^[5] the different root zone temperatures 164 did not significantly affect the SO²⁻⁴ concentration in the roots (Table 3). However, in 165 166 tubers, the RZT in T1 induced higher redistribution of this elements and the 167 temperatures generated by T2 and T3 reduced the SO²⁻⁴ concentration significantly, 168 possibly due to a high reduction and assimilation in the aerial part in organic compounds, 169 giving the low concentration in the tubers. With regard to the aerial part (Table 3), both 170 in stems and leaflets, the SO²⁻⁴ concentrations in T2 and T3 were the lowest. The reduction of SO²⁻₄ under this conditions was higher and the SO²⁻₄ which are highly mobile 171 in the xylem would be assimilated in the leaflets, while in T0 and T1, occured high levels 172 173 of SO²⁻₄ possibly for a decrease in assimilation.^[4]

174 The organic-S concentrations reflects S fraction in organic structures and represents its assimilation, varying similarly to the total-S.^[21] In roots, the highest 175 176 organic-S concentration was also recorded in T3 (26% higher than T0; Table 4) and the 177 lowest values were recorded in T0 and T1. In tubers, except T1, which gave the lowest 178 concentration (18% lower than T0), the rest of treatments did not statistically differ from 179 each other. In stems, non-significant differences were found between treatments. 180 Finally, in leaflets, T3 also gave the highest concentration (23% higher than T0), and no 181 significant differences were found between the rest of treatments.

With respect to this organic form of the S in the roots (Table 4), the temperatures induced by T2, T3 and T4 favoured a higher assimilation of SO²⁻₄, in comparison with the lower root temperatures of T0 and T1.^[22] In the tubers, except for the T1 treatment, with low concentrations of organic S, the temperatures generated by other treatments did not influence its redistribution. For the aerial part (Table 4), we observed that the 187 concentration of organic-S in the stems was not significantly affected by the root zone
188 temperatures, while in the leaflets T3 induced maximum concentration of this S forms,
189 implying a higher SO²⁻₄ reduction and a high transport rate of SO²⁻₄ from the roots,^[5]
190 while the lower temperatures induced by T0 and T1, and the too high temperatures of
191 the T4 had a negative influence on these processes.

The total-S in roots (Table 5), presented in T3 the highest concentration surpassing T0 by 22% and the lower values were found in T0 and T1, while in tubers, non-significant differences were found between treatments. For stems, the highest total-S concentrations were found in T0 and T4 and the lowest in intermediate treatments. In leaflets, T3 gave an increase of 19% with respect to T0.

197 Therefore, the different root-zone temperatures significantly influenced the total-198 S concentration (Table 5), with lower concentrations of total-S in the roots in T0 and T1 199 as a result of the lower root-zone temperatures,^[23] affecting significantly the S uptake 200 and assimilation as well as growth, in comparison with the treatments of high 201 temperatures, mainly the T3 that favoured a higher assimilation and concentration of 202 this macronutrient. In the tubers, the root-zone temperature did not affect the total-S 203 concentrations and for the stems, the highest levels were given in the treatments with 204 the highest (T4) and the lowest root temperatures (T0), since the root-zone temperature 205 directly affect the total-S concentration, because the effect was exerted on the 206 concentrations of SO²⁻⁴ and organic-S, while the range of more appropriate 207 temperatures, T2 and T3, reduced the concentration of total S in the stems. In the 208 leaflets (Table 5), in order to favour the synthesis of amino acids as tolerance mechanism 209 to toxic elements, would interest a higher growth and S concentrations,^[24] since a high 210 level of organic-S was observed in T3 (treatment with a high concentration of Se), a 211 higher SO²⁻₄ assimilation was favoured in the leaflets.

For plants, Se toxicity results primarily from the interference with the sulfur 212 metabolism,^[7] and most agricultural crops have low Se tolerance (<50mgSekg⁻¹ d.w.). In 213 relation to such phytotoxicity, according to Pais and Jones, [24] the normal Se content is 214 0.02mgkg⁻¹ leaf d.w., while in our experiments the plants accumulated quantities 215 exceeding 0.4–0.5mgkg⁻¹ without any toxicity symptoms. Ulrich,^[25] suggested that the 216 normal concentrations of S and SO²⁻₄ are between 0.8–3.0mgg⁻¹ leaf d.w., and between 217 0.25–1mgg⁻¹ leaf d.w., respectively. Our potato plants presented higher concentrations 218 219 in leaflets (25–30mg g⁻¹S d.w.; 3–4mgg⁻¹SO²⁻⁴ d.w.) probably as a tolerance mechanism 220 of these plants to the high S status.

According to Salt and Krämer,^[26] a plant is a hyperaccumulator if the ratio 221 222 (concentration of the metal in the aerial part):(concentration of metal in the root part) 223 exceeds 1. According to this, for Se, the ratio exceeded 1 and implies a potential for hyperaccumulation.^[27] Although the Se levels is below 0.1% d.w. in the aerial part, these 224 225 potato plants provides an advantage in phytoremediation against the techniques based on engineering, which are costly and also cause pollution.^[11] Therefore, there is a need 226 227 to improve the possibilities of accumulation of elements in potato crops and/or in other 228 species, using the mulch technique to ensure phytoextraction by manipulation of the 229 root-zone temperatures.

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231 ACKNOWLEDGEMENTS

The authors express their gratitude to the "Instituto Nacional de Investigación y Tecnología Agraria y Alimentaria" (INIA) and the "Dirección General de Investigación Agraria de la Consejería de Agricultura y Pesca de la Junta de Andalucía" for the financial

support for this work within the framework of Research Projects "INIA 8505" and "INIA 235 236 SC93-084" and to the C.I.F.A. (Centro de Investigación y Formación Agraria) for its 237 support in the experiments, and plant and soil sampling. The authors would like to thank 238 David Nesbitt for the translation into English, reviewing and constructive comments. 239 240 REFERENCES Engels, C.; Marschner, H. Effect of Suboptimal Root-Zone Temperature and Shoot **241** 1. 242 Demand on Net Translocation of Micronutrients from the Roots to the Shoot of 243 Maize. Plant Soil 1996, 186, 311–320. **244** 2. Manrique, L.A. Mulching in Potato Systems in the Tropics. J. Plant Nutri. 1995, 18, 245 593-616. **246** 3. Hanna, H.Y. Black Polyethylene Mulch does not Reduce Yield of Cucumbers Double 247 Cropped with Tomatoes under Heat Stress. HortScience 2000, 32, 190–191. 248 4. Leustek, T.; Saito, K. Sulfate Transport and Assimilation in Plants. Plant Physiol. 1999, 249 120, 637–643. **250** 5. Zhao, F.J.; Withers, P.J.A.; Evans, E.J.; Monaghan, J.; Salmon, S.E.; Shewry, P.R.; 251 McGrath, S.P. Sulphur Nutrition: An Important Factor for the Quality of Wheat and 252 Rapseed. Soil Sci. Plant Nutr. 1997, 43, 1137–1142. 253 6. Mayland, H.F.; James, L.F.; Penter, K.E.; Sonderegger, J.L. Selenium in Seleniferous Environments. In: Selenium in Agriculture and the Environments; Jacobs, L.W., Ed.; 254 255 American Society of Agronomy: Madison, WI, 1989; 15–50. **256** 7. Mikkelsen, R.L.; Page, A.L.; Bingham, F.T. Factors Affecting Selenium Accumulation by 257 Agricultural Crops. In: Selenium in Agriculture and the Environment. Jacobs, L.W., Ed.; 258 American Society of Agronomy: Madison, WI, 1989, 65–94. **259** 8. Schrauzer, G.N.; Sacher, J. Selenium in the Maintenance and Therapy of HIV Infected 260 Patients. Chem. Bio. Interactions. 1994, 91, 199–205. **261** 9. Bañuelos, G.S.; Mayland, H.F. Absorption and Distribution of Selenium in Animal 262 Consuming Canola Grown for Selenium Phytoremediation. Ecotoxicology and 263 Environmental Safety 2000, 46, 322–328. 264 10. Salt, D.E.; Smith, R.D.; Raskin, I. Phytoremediation. Ann. Rev. Plant Phyiol. Plant Mol. 265 Biol. 1998, 49, 643–668. 266 11. Meagher, R.B. Phytoremediation of Toxic Elemental and Organic Pollutants. Current 267 Opinion in Plant Biology 2000, 3, 153–162. 12. Jones, J.B. Jr. Plant Tissue Analysis in Micronutrients. In: Micronutrients in 268 269 Agriculture, 2nd Ed.; Mortvedt, J.J., Cox, F.R., Shuman, L.M., Welch, R.M., Eds.; SSSA 270 Book Serie No.4. S.S.S.A. Inc.: Madison, Wisconsin, USA, 1991; 477–521. 271 13. Novozamsky, I.; Van Eck, R. Total S Determination in Plant Material. Z. Anal. Chem. 272 1977, 286, 367–368. 273 14. Wolf, B.A. Comprehensive Systems of Leaf Analysis and Its Use for Diagnosing Crop 274 Nutrients Status. Commun. Soil Sci. Plant Anal. 1982, 13, 1035–1059. 275 15. Ham, J.M.; Kluitenberg, G.J.; Lamont, W.J. Optical Properties of Plastic Mulch Affect 276 the Field Temperature Regime. J. Amer. Soc. Hort. Sci. 1993, 118, 188–193

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	RZT	Roots	Tubers	Stems	Leaflets
		Biomass g Plant ¹			
Treatments	°C				
то	16 e ^a	1.75 bc	19.94 c	1.82 b	2.48 b
T1	20 d	1.04 c	10.89 d	2.19 a	1.93 c
Т2	23 c	1.97 b	22.42 b	1.85 b	2.59 b
Т3	27 b	2.34 a	26.93 a	1.70 c	2.84 a
T4	30 a	1.63 bc	20.70 c	2.09 ab	2.51 b

Table 1.Effect of Mulch Treatments on Root-Zone Temperature (RZT) and on Biomass (Dry Weight) of Potato Organs

	Roots	Tubers	Stems	Leaflets
Treatments		Mgg⁻ੰ	^L d.w.	
т0	209 cd ^a	_Y	448 b	263 c
T1	115 d	-	465 ab	206 d
Т2	225 c	-	154 d	322 b
Т3	315 a	-	195 c	423 a
T4	260 b	-	489 a	307 bc

Table 2. Effect of Root-Zone Temperatures on Total Se Concentration in Potato Organs

^aMean values followed by the same letter within a column were not significantly different at p<0.05 according to Duncan's Multiple Range Test. ^yConcentration below detection limits.

	Roots	Tubers	Stems	Leaflets
Treatments	Mgg ⁻¹ d.w.			
т0	3.61 a ^a	2.45 b	4.05 a	3.38 ab
T1	3.73 a	3.84 a	4.31 a	3.89 a
Т2	3.72 a	1.66 c	3.14 b	3.18 b
Т3	3.65 a	1.61 c	3 b	3.07 b
T4	3.60 a	2.49 b	3.01 b	3.41 ab

Table 3. Effect of Root-Zone Temperatures on SO²₄ Concentration in Potato Organs

	Roots	Tubers	Stems	Leaflets
Treatments		mgg ¹	d.w.	
т0	20.98 b ^a	13.24 a	25.47 a	22.76 b
T1	20.49 b	10.82 b	22.11 a	21.28 b
Т2	23.57 ab	13.96 a	23.11 a	23.53 b
Т3	26.48 a	14.28 a	23.67 a	28.11 a
T4	23.47 ab	13.56 a	26.46 a	22.67 b

Table 4. Effect of Root-Zone Temperatures on Organic-S Concentration in Potato Organs

	Roots	Tubers	Stems	Leaflets
Treatments	mgg ¹ d.w.			
т0	24.6 c ^a	15.7 a	29.5 a	26.1 b
T1	24.2 c	14.7 a	26.4 b	25.2 b
Т2	27.2 b	15.6 a	26.3 b	26.7 b
Т3	30.1 a	15.9 a	26.7 b	31.2 a
T4	27.1 b	16.1 a	29.5 a	26.1 b

Table 5. Effect of Root-Zone Temperatures on Total-S Concentration in Potato Organs